Original Article

Outcomes of Anti-CD19 CAR-T Treatment of Pediatric B-ALL with Bone Marrow and Extramedullary Relapse

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Running Title: CD19 CAR-T for B-ALL with BM and EM Relapse

This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process which may lead to differences between this version and the Version of Record. Please cite this article as an 'Accepted Article', doi:10.4143/crt.2021.399
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Abstract

Purpose
Anti-CD19 chimeric antigen receptor T-cell immunotherapy (19CAR-T) has achieved impressive clinical results in adult and pediatric relapsed/refractory (r/r) B-lineage acute lymphoblastic leukemia (B-ALL). However, the application and effect of CAR-T therapy in B-ALL patients with extramedullary relapse are rarely issued even disqualified in some clinical trials. Here, we examined the efficacy of 19CAR-T in patients with both bone marrow and extramedullary involvement.

Materials and Methods
CAR-T cells were generated by transfection of primary human T lymphocytes with a lentiviral vector expressing anti-CD19 single chain antibody fragments (scFvs) with the cytoplasmic domains of 4-1BB and CD3ζ, and used to infuse patients diagnosed as having r/r B-ALL with extramedullary origination. Clinical responses were evaluated by the use of bone marrow aspiration, imaging, and flow cytometry.

Results
Eight patients received 19CAR-T infusion and all attained complete remission (CR). Only one patient was bridged to hematopoietic stem cell transplantation (HSCT). Although three patients relapsed after infusion, they received 19/22CAR-T infusion sequentially and attained a second remission. To date, five patients are in continuous CR and all eight patients are still alive. The mean follow-up time was 21.9 months, while the 24-month estimated event-free survival is 51.4%.

Conclusion
Anti-CD19 CAR-T therapy can lead to clinical remission for extramedullary relapsed pediatric B-ALL patients. However, the problem of CD19+ relapses after CAR-T remained to be solved. For patients relapsing after CAR-T, a second CAR-T therapy creates another opportunity for remission for subsequent HSCT.

Keywords
B-ALL, extramedullary relapse, CD19 CAR-T, second CAR-T
Introduction

Acute lymphoblastic leukemia (ALL) is a leading hematological malignancy of children worldwide. Although the long-term survival rate for pediatric ALL approaches and even exceeds 90%, the 10–15% of cases of ALL that relapse are considered one of the leading causes of cancer mortality in children [1,2]. Although relapse is mainly detected in bone marrow, occasionally extramedullary tissues are involved and these account for 15–20% of all relapses [3]. Extramedullary relapse is most commonly identified as central nervous system leukemia or testicular leukemia [4,5]. The natural history of extramedullary relapse generally correlates with poor prognosis and limited overall survival [6]. Conventional chemotherapy, radiation, or even allogeneic hematopoietic stem cell transplantation (allo-HSCT) tend to be unsuccessful in these patients, especially those with multiple relapses [7,8]. Therefore, the treatment of ALL patients with extramedullary disease remains a great challenge and new therapeutic strategies are urgently required.

Recently, chimeric antigen receptor T-cell (CAR-T) immunotherapy has been receiving wide attention and becoming an increasingly hot topic in the treatment of relapsed/refractory (r/r) B-lineage acute lymphoblastic leukemia (B-ALL). CAR-T cells were originally developed by Eshhar and colleagues in the late 1980s and early 1990s [9]. The approach harnesses the cytotoxicity of genetically modified T cells with high specificity for tumor cells. CAR-T cells contain an antigen-specific single-chain antibody fragment (scFv) linked to intracellular signaling domains independent of MHC restriction and antigen processing [10]. There is no doubt that CD19-targeted CAR-T therapy (19CAR-T) has achieved impressive clinical results in both adult and pediatric r/r B-ALL, with an amazing disease remission rate, and is emerging as a standard strategy for r/r B-ALL patients [11-14]. Nevertheless, the application and effect of CAR-T therapy in B-ALL patients with extramedullary relapses has rarely been described
and such patients are sometimes excluded from some clinical trials [11,13].

Here we report our experiences using anti-CD19 CAR-T cells to treat 8 pediatric B-ALL patients with simultaneous extramedullary and bone marrow relapse.

Methods

1. Study design

Between September 2017 and July 2020, 8 pediatric patients with CD19+ B-ALL relapsed with both bone marrow and extramedullary sites and were enrolled in this study. The study was conducted in Shanghai Children’s Medical Center, which is affiliated to Shanghai Jiao Tong University School of Medicine and Children’s Hospital of Soochow University, and was registered at chictr.org.cn (ChiCTR2100041852) from November 2016. Informed consent forms were signed by the patients’ guardians and were approved by the ethics committee of Shanghai Children’s Medical Center and Children’s Hospital of Soochow University.

2. Cell Production

Peripheral blood (PB) was drawn directly from patients for T-cell harvesting using anti-CD3-coated beads (Invitrogen). For patients who relapsed after HSCT, PB from a donor was used. Anti-CD19 scFvs derived from FMC63 were ligated into a lentiviral vector, downstream of an EF1α promoter and in a backbone with the CD8A signal peptide, hinge, CD8 transmembrane domains, and the cytoplasmic domains of 4-1BB and CD3ζ to create a second-generation CAR construct. The manufacture of CAR-T cells usually took 7–10 days. Prior to CAR-T injection, all patients were given lymphodepleting chemotherapy: 500 mg/m² cyclophosphamide for 2 days (days −4 and −3) and 40 mg/m² fludarabine for 3 days (days −4, −3, and −2). On day 0, patients were infused with fresh CAR-T cells at a dosage of 1–15 × 10^6.
10^6/kg. Patients who relapsed after the first exposure to CAR-T cells were allowed to receive a second CAR-T, and given 500 mg/m² cyclophosphamide and 50 mg/m² fludarabine per day for 3 days (days –4, –3, and –2) for lymphodepletion. For a combined CAR-T therapy, we combined anti-CD19 CAR-T cells with anti-CD22 CAR-T cells whose scFv was derived from the 5/44 clone.

3. Patient Evaluation

Before CAR-T administration, tumor burden was examined by flow cytometry of bone marrow, cerebrospinal fluid (CSF), or biopsy samples. Neurologic adverse events were graded according to the Common Terminology Criteria for Adverse Events (CTCAE) 4.03, and cytokine release syndrome (CRS) was graded as per the American Society for Transplant and Cellular Therapy grading criteria (ASTCT) [15]. A bone marrow aspiration was done 7–14 days after CAR-T infusion, and disease burden assessment was performed on marrow samples. Disease-free marrow was defined as having less than 0.01% abnormal cells identified by multiparameter flow cytometry performed at the Key Laboratory of Hematology and Oncology Ministry of Health. Additionally, complete remission (CR) status was defined as no leukemic blasts detected in both bone marrow (BM) and extramedullary sites. For central nervous system (CNS)-involved patients, CSF was collected for analysis. For patients with subcutaneous and/or testicular tumors, changes in tumor size were observed and compared, and biopsies were performed for flow cytometry. Examinations of the relevant extramedullary sites were performed on days 7 and 14, and at 1 month, 2 months, 3 months, 6 months, 1 year, and 2 years following CAR-T cell infusion. CAR-T cell persistence in the PB or CSF of patients was measured using real-time qPCR and copies per ug DNA were normalized to the single-copy gene CDKN1a.
4. Cytokine Analysis

Quantification of PB and CSF soluble cytokines including IL2, IL4, IL6, IL10, TNFα, IFNγ, and IL17 was performed using a BD Cytometric Bead Array Human Th1/Th2/Th17 Cytokine Kit, according to the manufacturer’s instructions.

5. Statistical Analysis

For event-free survival (EFS), an event was defined as bridging to HSCT, or relapse, or death. Death was the event for analysis of overall survival (OS). Kaplan-Meier curves were created for EFS and OS analysis and compared using the log-rank test. All statistics were performed as indicated, using Graph-Pad Prism 8 software for Windows, version 8.0.1. Statistically significant differences were defined at a p-value <0.05.

Results

Patient characteristics are shown in Table 1. Briefly, a total of eight extramedullary relapsed childhood B-ALL patients with an average age of 7.9 years (range, 4.3–13.0) were enrolled in this study and treated at Shanghai Children’s Medical Center (six patients) and Children’s Hospital of Soochow University (two patients). There were four patients whose cancer infiltrated the CNS, one with a testis tumor, two with a subcutaneous tumor, and one with both CNS and testis involvement. All patients’ disease had relapsed. The mean follow-up time was 21.9 months (5.93–39.97 months). In addition, five of eight patients (39.4%) relapsed more than once, of whom only patient S003 relapsed after HSCT. Therefore, S003 was given donor-derived CAR-T cells from her 7/10 HLA-matched father, without developing GVHD. All patients received fresh CAR-T product and the average infusion dose was $9.9 \times 10^6$ per kilogram of body weight (range, 1.8–15.0 $\times 10^6$).
Patient-level clinical response outcomes are exhibited in Fig. 1A. All eight patients achieved CR: 5 patients are now in continuous CR, and no CAR-T-related death occurred. Patient time to CR was usually 2–6 weeks. To date, only one patient (S006) chose bridging to HSCT after the second CAR-T. The estimated EFS of patients with 19CAR-T cells was approximately 68.57% at 12 months and decreased to 51.43% at 24 months (Fig. 1B).

Except patient S006, CRS was observed in all patients, and it was above grade 2 in three of eight patients. CAR-T related encephalopathy syndrome (CRES) was observed in five patients (62.5%), with no CRES above grade 2 identified. Four patients received tocilizumab, and two of these patients simultaneously received glucocorticoids. Only patient S004 received glucocorticoids alone, by intrathecal injection. No patient died of severe CRS or CRES (Table 1). Interleukin 6 (IL-6) and interferon γ (IFNγ) concentrations are the main biomarkers of CRS severity. Similar patterns were observed in both IL-6 and IFNγ (Fig. 2A); the highest level of IL-6 was 60618.45 pg/mL, detected in patient S003 (Table 1). On day 7, transient increases in levels of serum IL-6 and IFNγ occurred in most patients during CRS after CAR-T administration. We also observed that a drop of IL-6 was always followed by an increase in lymphocyte numbers in the PB (Fig. 2B) that was temporally coincident with the resolution of CRS.

In four patients with subcutaneous or testicular infiltration, CRS initially presented as a painless soft swelling of the affected sites along with redness and fever, which usually would last for about 3 days after CAR-T infusion. After a peak in swelling at approximately day 7, the tissue would return to normal size. Finally, the clearance of leukemic cells in the site was confirmed in all by flow cytometry of cells from biopsies and BM.

Patients with CNS involvement had more severe symptoms of encephalopathy, manifested as high fever, headache, frequent vomiting, drowsiness, unconsciousness, hypotension or hypertension, limb trembling, and convulsions (Table 1). For the patient with convulsions,
lumber puncture was required, and the CSF protein increased significantly; meanwhile, the CSF IL-6 concentration was much higher than that in PB. For example, on day 6, patient S004 suffered 1272.27 pg/mL of IL-6 in the CSF but 26.62 pg/mL in the PB. We also detected a higher level of the CAR gene in CSF compared with PB. In patient S005, 465.026 × 10⁳ copies of CAR per ug DNA were detected, over 10-fold the number detected in PB on day 3 (Fig. 2C). Additionally, CAR-T cell expansion determined using real-time qPCR revealed that the peak CAR-T expansion was variable in different extramedullary-relapsed patients but mostly not observed after Day 7 (Fig. 2D). About 2–3 weeks later, the absence of remaining leukemic cells was verified by flow cytometry of both bone marrow and CSF samples (Fig. 2F). During follow-up, the longest period of B-cell aplasia was found in patient S003, extending more than 200 days after CAR-T cell infusion; this was also indirectly confirmed using qPCR and Sanger sequencing for the existence of CAR-T cells (Fig. 2E).

Patients S002, S004, and S006 had relapsed after prior exposure to CAR-T cells. Their characteristics and responses are presented in Table 2. Patient S006 relapsed with an isolated subcutaneous tumor, while the other two patients had isolated BM relapses. Due to the high expression of CD19 and CD22 on the relapsed leukemic blasts, these patients accepted a combined CD19- and CD22-targeting second patient-derived CAR-T cell therapy. All of them then achieved CR again. All three patients experienced only grade 1 CRS along with fever. In patient S006, 23.45 × 10³ copies of CAR per ug DNA were detected on day 89 after the second CAR-T infusion, which was higher than the 14.93 copies of CAR per ug DNA on day 60 after her first 19CAR-T therapy. B-cell asplasia was observed in patient S002 on day 7 after CAR-T infusion. However, she rejected bridging to HSCT and then relapsed again after 9.3 months. At her parents’ strong insistence, a third combined CAR-T therapy was conducted because of the remaining CD19- and CD22-expressing leukemic cells.
Discussion

There is no optimal defined protocol for treating extramedullary leukemia. Some patients experience multiple relapses even after receiving chemotherapy, allo-HSCT, and/or radiation [16, 17]. Before off-the-shelf CAR-T therapy, blinatumomab had received regulatory approval and was widely available to treat relapsed or refractory B-cell precursor ALL patients [18]. However, extramedullary relapse is associated with a lower CR rate in patients treated with blinatumomab, which is a CD19/CD3 bispecific T-cell engager antibody that induces the death of CD19+ leukemia cells by redirecting CD3+ T cells [19]. Furthermore, many patients in whom blinatumomab therapy has failed still have CD19+ leukemic cells [20]. Two studies have reported that CD19 CAR-T therapy can eradicate leukemic cells in B-ALL patients with extramedullary involvement. In a study of 7 boys with testicular relapse, 6 of them remained in CR for a median of 14 months (5–23 months) [21]. The group from the Children's Hospital of Philadelphia demonstrated that 6 in 10 patients with various extramedullary sites of relapse were without recurrence of disease for 6–13 months after CAR-T infusion [22]. A clinical remission in extramedullary B-ALL patients with anti-CD19 CAR-T therapy was also noted in that study.

Extramedullary ALL presents a challenge similar to that of solid malignancies: the trafficking of targeted cells to the tumor sites. Some studies have found that CAR-T cells may migrate into and lead to partial tumor regression in extramedullary sites such as liver, lung, etc. [16,23,24]. However, the effect on testicular or CNS ALL remains unclear. For example, the absence of detectable donor cells in extramedullary lesions has been reported [25]. This suggests that intravenously injected CAR-T cells may not reach isolated extramedullary sites, and thus may have no effect. Intriguingly, the high rates of CR achieved with intravenous administration in this cohort are evidence that CAR-T cells can traffic to extramedullary sites.
and then eliminate tumor cells efficiently. However, there was a high incidence of immune effector cell–associated neurotoxicity syndrome, especially in B-ALL relapsed patients with CNS involvement. This may be associated with increases in CSF IL-6 levels (Table 1), which are thought to not be prevented by the blood-brain barrier [26]. The expression of IL-6 has been reported to be related to endothelial activation and blood-brain barrier dysfunction after CD19 CAR-T infusion [27].

The value of consolidative BMT after CAR-T therapy remains controversial. On the one hand, Lee et al. have suggested that BMT following the infusion of CAR-T cells enhances the survival of children and young adult patients [28]. On the other hand, Park et al. found that the survival improvement was not significant in adult patients [29]. The costimulatory domain in the above two studies was CD28. Interestingly, a recent study showed that allo-HSCT achieved better EFS in patients aged over 18 years after treatment with CAR-T cells incorporating a 4-1BB costimulatory domain [30]. In this study, only patient S001, who was one of five non-relapsed patients, was followed with HSCT, but he gained the longest CR.

We note that 100% CR was achieved in the three relapsed patients in our study. This indicates that retreatment with CAR-T cell infusion may offer an opportunity for B-ALL pediatric patients with extramedullary relapse to wait and look for a bridge to effective HSCT. This result also suggests that combining CAR-T cells that target different antigens can be an effective solution to this kind of immune escape problem, which is associated with tumor-cell antigen switching.

This study reveals that anti-CD19 CAR-T therapy can partially induce clinical remission in patients with both bone marrow and extramedullary relapses, while CD19+ relapse remains the main problem. We suggest a second CAR-T cell treatment as a means to create another chance for survival in these patients.
Ethical Statement

The study was approved by the IRB of Shanghai Children's Medical Center affiliated to the Medical College of Shanghai Jiaotong University (IRB number: SCMCIRB-K2016067-1). Written informed consent for anti-CD19 CAR-T therapy was obtained from patients’ guardians.

Author Contributions

Conceived and designed the analysis: Wan X, Chen J, Tang Y, Lu J, Li B.


Performed the analysis: Wan X, Yang X, Wang T, Li B.

Wrote the paper: Wan X, Yang X, Li B.

Conflicts of Interest

Conflict of interest relevant to this article was not reported.

Acknowledgements

This study was supported by the Shanghai Collaborative Innovation Center for Translational Medicine (No. TM201928) and the National Natural Science Foundation of China (81670174 to B. L, 81900158 to L. S).
References


<table>
<thead>
<tr>
<th>Patient ID</th>
<th>Sex</th>
<th>Age (years)</th>
<th>Tissue Involvement</th>
<th>Enrollment Criteria</th>
<th>Time from Initial Diagnosis to CAR-T infusion, months</th>
<th>Genetic variation</th>
<th>Minimal Residual Disease before CAR-T (%)</th>
<th>Infusion Dose of 19CAR-T ($\times 10^6$/kg)</th>
<th>CRS Grade</th>
<th>CRES Grade</th>
<th>IL-6 max (pg/mL)</th>
<th>CNS Status at the time of CAR-T cell infusion</th>
<th>Disease Status on day 28*</th>
</tr>
</thead>
<tbody>
<tr>
<td>S001</td>
<td>M</td>
<td>4.3</td>
<td>BM, subcutaneous tumor</td>
<td>Relapse</td>
<td>7.50</td>
<td>TCF3-PBX1</td>
<td>95.83</td>
<td>6.9</td>
<td>1‡</td>
<td>0</td>
<td>390.25</td>
<td>ND</td>
<td>MRD Neg, 30 $\times 21$mm*</td>
</tr>
<tr>
<td>S002</td>
<td>F</td>
<td>7.2</td>
<td>BM and CNS</td>
<td>2nd Relapse</td>
<td>44.07</td>
<td>-</td>
<td>0.27</td>
<td>10</td>
<td>1</td>
<td>1</td>
<td>100.39</td>
<td>ND</td>
<td>MRD Neg</td>
</tr>
<tr>
<td>S003</td>
<td>F</td>
<td>6.1</td>
<td>BM and CNS</td>
<td>2nd Relapse after HSCT</td>
<td>50.10</td>
<td>ETV6-RUNX1</td>
<td>31.35</td>
<td>12</td>
<td>3‡</td>
<td>0</td>
<td>60618.45</td>
<td>ND</td>
<td>MRD Neg</td>
</tr>
<tr>
<td>S004</td>
<td>F</td>
<td>8.7</td>
<td>BM and CNS</td>
<td>2nd Relapse</td>
<td>72.87</td>
<td>ZCCHC7-CSF2RA</td>
<td>2.41</td>
<td>15</td>
<td>1‡</td>
<td>1</td>
<td>1272.27†</td>
<td>Mild Headache Drowsiness and slow response to stimuli</td>
<td>MRD Neg</td>
</tr>
<tr>
<td>S005</td>
<td>M</td>
<td>9.3</td>
<td>BM and testis</td>
<td>Relapse</td>
<td>40.10</td>
<td>ETV6-RUNX1</td>
<td>1.00</td>
<td>15</td>
<td>3‡</td>
<td>2</td>
<td>1445.5</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>S006</td>
<td>F</td>
<td>7.4</td>
<td>BM, subcutaneous tumor</td>
<td>Relapse</td>
<td>44.17</td>
<td>ND</td>
<td>0.46</td>
<td>10</td>
<td>0</td>
<td>0</td>
<td>96.4</td>
<td>ND</td>
<td>-</td>
</tr>
<tr>
<td>S007</td>
<td>M</td>
<td>7.1</td>
<td>BM, CNS, and testis</td>
<td>2nd Relapse</td>
<td>48.27</td>
<td>-</td>
<td>3.00</td>
<td>8.8</td>
<td>2‡</td>
<td>2</td>
<td>13116.22</td>
<td>Limb trembling and delirium Unconsciousness</td>
<td>MRD Neg</td>
</tr>
<tr>
<td>S008</td>
<td>F</td>
<td>13</td>
<td>BM and CNS</td>
<td>2nd Relapse</td>
<td>116.83</td>
<td>ETV6: exon 2 del</td>
<td>32.40</td>
<td>1.8</td>
<td>2‡</td>
<td>2</td>
<td>574.7†</td>
<td>-</td>
<td>-</td>
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</table>

M, male; F, female. ‡Tocilizumab. §Corticosteroid. ‡CSF level. ND, not detected; -, no follow-up. CRS, cytokine release syndrome; CRES, CAR-T related encephalopathy syndrome. S004 received glucocorticoids by intrathecal injection. *MRD neg, undetectable BM and CNS MRD (minimal residual disease) as measured by flow cytometry. **According to the results of ultrasound, patient S001 was diagnosed with a 45 $\times$ 25 mm enlarged lymph node near an iliac vessel in the pelvis before CAR-T infusion; the dimensions decreased to 30 $\times$ 21 mm after 28 days, and after 3 months, only one 18 $\times$ 10 mm lymph node was found in the rightinguinal region.
Table 2. Patients with a Second Relapse After 19CAR-T.

<table>
<thead>
<tr>
<th>Patient ID</th>
<th>2nd Relapse Sites</th>
<th>CD19/CD22 Expression after 2nd Relapse (%)</th>
<th>Infusion Dose of 19/22CAR-T (×10⁶/kg)</th>
<th>19CAR-T in All (%)</th>
<th>CRS Grade</th>
<th>CRES Grade</th>
<th>IL-6 max (pg/mL)</th>
<th>Responses</th>
<th>B cell on Day 7 after CAR-T infusion (cells/μL)</th>
<th>Bridge to HSCT</th>
</tr>
</thead>
<tbody>
<tr>
<td>S002</td>
<td>BM</td>
<td>-</td>
<td>15</td>
<td>60.0</td>
<td>1</td>
<td>0</td>
<td>24.17</td>
<td>CR*</td>
<td>0.0</td>
<td>N</td>
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<tr>
<td>S004</td>
<td>BM</td>
<td>98.3/99.7</td>
<td>4.26</td>
<td>51.3</td>
<td>1</td>
<td>0</td>
<td>-</td>
<td>CR</td>
<td>-</td>
<td>N</td>
</tr>
<tr>
<td>S006</td>
<td>Subcutaneous tumor</td>
<td>100/100</td>
<td>12</td>
<td>20.6</td>
<td>1</td>
<td>0</td>
<td>-</td>
<td>CR</td>
<td>-</td>
<td>Y</td>
</tr>
</tbody>
</table>

-, no follow-up; CRS, cytokine release syndrome; CRES, CAR-T related encephalopathy syndrome; CR, complete remission. *Third relapsed after 2nd CAR-T infusion.
Fig. 1. Clinical outcomes of anti-CD19 CAR-T cells. (A) Swimmer plot (n = 8), in which each bar represents an individual patient as designated. (B) Kaplan-Meier graph of EFS in 8 patients infused with 19CAR-T cells, demonstrating 68.57% event-free survival at 12 months and 51.43% at 24 months.
Fig. 2. Changes in patients’ serum biomarkers in CRS. (A) The temporal relationship of IL-6 (red line) and IFNγ (blue line) from day 0 to day 11. (B) The temporal relationship of IL-6 (red line) and lymphocyte numbers (blue line) from day 0 to day 8; L: lymphocytes. (C) The number of copies of the CAR gene detected in Patient S005’s PB and CSF; PB, peripheral blood; CSF, cerebrospinal fluid. (D) The persistence of circulating CAR-T cells identified by qPCR. (E) B-cell aplasia remained in patient S003 but not in S001, who had been bridged to HSCT. (F) There were 90.7% leukemic cells in BM from patient S003 detected on day 0 (left) but 0.0% on day 21 (right).